

# Comparative Analysis of Plants-Based Materials as Natural Coagulant in Raw Water Treatment

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## ABSTRACT

The research was carried out to compare the potentials and effectiveness of plant-based materials as coagulants in raw water treatment. Aloe Vera Gel, Seeds of Moringa, Tamarind and Water Melon were used as natural coagulants to treat raw water. Moringa seed powder, Aloe Vera gel and Tamarind seeds powder and Aloe Vera gel and Water Melon Seeds Powder were prepared. Jartest analysis was carried out, using stirring time of 30 minutes and 400 rpm throughout the experiment. Settling time, pH, turbidity removal, percentage of coagulation performance, TDS and TSS were analyzed. Settling time of 5 minutes, pH of 7.21, final turbidity of 21 coagulation performance of 95.3%, turbidity removal of 429 NTU, TDS of 118 mg/l and TSS of 0.08 g were recorded for Moringa seed powder. Also, 4 minutes, 18 NTU, 6.93, 96%, 432 NTU, 138 mg/l and 0.08 g were recorded using Aloe vera gel for settling time, final turbidity, pH, percentage of coagulation performance, turbidity removal, TDS and TSS respectively. Also, 23 minutes, 7.31, 33 NTU, 92.5%, 417 NTU, 160 mg/l and 0.32 g were recorded using Tamarind seeds powder for settling time, pH, final turbidity, percentage of coagulation performance, turbidity removal, TDS and TSS respectively. And 34 minutes, 6.97, 49 NTU, 91.1%, 138 NTU, 138 mg/L and 0.23 g were recorded using water melon seeds powder for settling time, pH, final turbidity, percentage of coagulation performance, turbidity removal, TDS and TSS respectively. It was discovered that all the plants-based materials (Aloe Vera, Moringa, Tamarind and Water Melon seeds) have the potential to effectively serve as coagulants and also have the potential to serve as disinfectants in raw water treatment.

**Keywords:** Aloe Vera gel, Moringa seeds, Tamarind seeds, Water Melon Seeds, coagulation performance, turbidity removal, Settling time and Stirring time.

## I. INTRODUCTION

Water is one of the basic needs for living organisms (both plants and animals) to live on Earth. For human beings, it is not water per se, but clean and pure one. Clean and pure water is the one which does not have any disease-causing pathogens and which undergoes proper water treatment processes, and thus ready for consumption (WHO, 2017). It is vital for societies to have clean and pure water, as shortage of it causes waterborne diseases, which are killer diseases (WHO, 2017). Access to sufficient amounts of clean and pure water is vital for human health than socio-economic development, because 70% of human body is made up of water (Kulinkina et al., 2016). That is why, World Health Organization (WHO, 2017), puts much emphasis on clean water and is always ensuring that people in both rural and urban areas have access to clean water; not only WHO, but government and private organizations. Despite the fact that 70% of the earth is covered with water, only 3% of the water is clean, pure and safe for drinking (WHO, 2011). These statistics, is what makes providing clean water to societies by policymakers daunting challenge, because currently about 1.1 billion people (both rural and urban) are facing and experiencing the problem of clean water shortage (WHO, 2011). This roughly affects 27% of world population and thus highlighting the real threat the problem is causing, which in the last decade is described as "Global Water Crisis" (Cain and Gleick, 2005) and "Global Water Scarcity" (United Nations, 2013). Failure to provide safe drinking water and adequate sanitation services to all people is perhaps the greatest development failure of the twentieth century (Gleick, 2005).

Inorganic coagulants refer to metal coagulants which are generally based on aluminum and iron. They are used in water and wastewater treatment. Due to their availability and effectiveness, they are the most used coagulant. They include: Aluminum Sulphate, Aluminum

Chloride, Aluminum Chlorohydrate, Sodium Aluminate, Polyaluminum Chloride, Polyaluminum Sulphate (Bratby, 2016). Iron coagulant include: Ferric Sulphate, Ferric Chloride, Ferrous Sulphate (Bratby, 2016). Hydrated lime and magnesium carbonate are other examples of metal coagulant, not based on aluminum and iron. Due to their effectiveness and availability.

Natural coagulants are derived from plants and animal source and are generally regarded as organic polymers. Natural coagulants are toxic free, biodegradable, cheap and available. It was reported that, over 50 natural coagulants are being reported for water treatment (Saleem and Bachman, 2019). Natural coagulants gain advantage over chemical coagulants, due to various reasons. Among them is that, natural coagulants are safer than chemical coagulants. By using chemical coagulants, there will be a possibility of residue present in the water after treatment, when consumed, the residue of chemical coagulants such as alum is harmful, by causing Alzheimer's disease (Garde and Buchberger, 2017). On the other hand, residue of natural coagulants is not harmful. Natural coagulants are cheaper compared to chemical coagulants. This is because during treatment, chemical coagulants like alum needs coagulant aid to effectively treat high turbid water, thus making it more expensive and difficult to be used in poor countries, whereas natural coagulants can easily be extracted from various plant wastes which reduce the cost of treatment (Antov et al., 2010). However, for natural coagulants to be used at commercial scale, it must be abundant and locally available. Natural coagulants have some disadvantages, because it increases the organic matter present, thus increasing microbial activity. Natural coagulants need longer sedimentation and settling time and the removal efficiency of bacteria is not high, thus bringing the possibilities of secondary bacterial growth (Awad et al., 2013).

### **1.1 STATEMENT OF PROBLEM**

The uses of inorganic and synthetic coagulants for water treatment is costly and have side effect to health of people, and not everyone have access to them. Recent studies have vindicated a number of serious drawbacks linked to the use of aluminum salts such as Alzheimer's disease associated with high aluminum residuals in treated water, excessive sludge production during water treatment and considerable changes in water chemistry due to reactions with the OH<sup>-</sup> and alkaliinity of water. In addition, the use of alum salts is inappropriate in some developing countries because of the high cost of imported chemicals and low availability of chemical coagulants (Adejumo et al., 2013). Also, most numbers of some synthetic organic polymers such as acrylamide have neurotoxicity and strong carcinogenic properties and because of this, there has been considerable interest in the development of natural coagulants which have

resafe for human health and biodegradable (Ghebremichael, 2004).

On the other hand, Aloe Vera, seeds of Moringa, Tamarind and Water Melon are abundant, cheap, and are always discarded. This is always adds to environmental pollution and their underutilization.

### **1.2 JUSTIFICATION**

Clean and Portable water is one of the major ingredients used in the production of most foods, especially soft drinks. Coagulation process is one of the main stages for the production of clean and portable water from surface water (raw water). Aloe Vera gel, seeds of Moringa, Tamarind and Water Melon serves as cheapest, available, biodegradable and easiest source of natural coagulants for water treatment, especially at domestic level. Harnessing the potential of these natural products will help in reducing environmental pollution, which they bring and producing a cheap and effective coagulant. People living in rural areas, use natural plant materials for household water treatment, thus, assessing and evaluating the potential of plant materials as natural coagulants in advanced water treatment technology will provide better and sustainable knowledge for improved household water treatment which can also be transferred back to rural areas.

### **1.3 BROAD OBJECTIVES**

The broad objectives of this research is to compare the potentials and effectiveness of plant-based materials as natural coagulants in raw water treatment

### **1.4 SPECIFIC OBJECTIVES**

The specific objectives of this work are:

To carry out jar test analysis for Aloe Vera gel, Moringa, Tamarind and Water Melon seed powder, and compare their effectiveness.

To analyze the physical parameters of the water after the jar test (pH, turbidity removal, coagulation performance, TDS and rate of settling).

To analyze the chemical parameters of the water after the jar test ( acidity, alkalinity, hardness and chlorides )

## **II. MATERIALS AND METHODS**

### **2.1 Sources of Materials**

The raw water sample was collected at the back of Zobe water treatment plants sample collection point. These seeds of the Moringa was obtained from a nearby irrigation farm at Dutsin-Matown; the Aloe Vera was obtained at the nursery garden in Dutsin-Matown, Katsina State Nigeria, while watermelon seed was obtained at Dutsin-Mama market

### **2.2.1 Preparation of Aloe Vera Gel**

The samples of the fresh Aloe Vera (25–30 cm) long was selected and collected in a polyethylene bag, and taken to the laboratory in the Zobe Water Treatment Plant. The Aloe Vera was washed thoroughly with clean water and cut into pieces using a razor blade. The pieces of the Aloe Vera were then ground using an electric blending machine. The blend was filtered to separate the gel from the fiber of the Aloe Vera. The gel was then introduced into the 500 ml distilled water and stirred well using a stirrer for 40 minutes. The solution was strained through a sieve of 30 mm. The filtrate was collected as Aloe Vera gel and stored in a refrigerator (Amruta et al., 2017).

### **2.2.2 Preparation of Moringa Seeds Powder**

The seeds of the moringa was washed with clean water, and then dried using oven at 55 degrees Celsius for 4 hours. The dried seeds will be ground using pestle and mortar, and the powdered Moringa seed will be obtained (Saleem and Buchman, 2019).

### **2.2.3 Preparation of Watermelon seed Powder**

The watermelon fruits was sliced open using a clean stainless-steel laboratory knife to obtain the seed. The seed was washed severally with water, sun-dried for a week, sorted to remove bad ones, shelled and ground with a high-speed laboratory electric blender, packed in a nail tight container. 50g of the crushed seeds was packed in a thin plastic bag and placed in a Soxhlet extraction apparatus. 300ml of n-

Hexane was used to extract oil from the crushed seed in the column. The apparatus was left running for about 5 hrs and stopped when the extraction was complete. The cake was washed with distilled water to remove residue of the n-Hexane, dried in an oven till constant weight and sieved. The finer particles was used as the coagulant (Eze et al., 2017).

### **2.2.4 Preparation of Tamarind Seeds Powder**

The whole seeds of the tamarind was washed severally with water, sun-dried for a week, sorted to remove bad ones, shelled and ground to a fine powder using a laboratory mill. All the rounded materials was sieved through 750 µm sieve and

he fraction with particle sizes less than 750 µm was used in experiments.

### **2.2.5 Procedure for Jar Test Analysis**

The jar test analysis was carried out by putting various grams (1g, 2g, 3g, 4g, 5g and 6g) of Aloe Vera gel in the six beakers containing the raw water, while the speed of them achiene was adjusted slowly. After varying the gramsoft he Aloe Vera gel, the speed was increased. The mixture of both the raw water and the gel was stirred at a speed of 400 rpm (revolution per minute), for 30 minutes. The jar machine was then switched off. The same procedure was repeated for Moringa seed powder, Tamarind seeds powder and Water Melon Seeds Powder respectively (APHA, 2005). The following observations were made during the analysis:

The rate of formation of flocs

The rate of settling of flocs

Clarity of the water

**2.2.6 Determination of pH:** Procedure of (APHA, 2005) was used

**2.2.7 Determination of Turbidity:** Procedure of (APHA, 2005) was used

**2.2.8 Determination of TDS:** Procedure of (APHA 2005) was used

**2.2.9 Analytical Methods**

The effects of these powdered seeds and gel as coagulant was determined based on the performance of coagulation activity and turbidity removal. Coagulation activity was conducted based on (Kukic et al., 2010) while turbidity removal expressed in NTU (nephelometric turbidity unit) was determined using turbidity meter HACH 2100 Q. Equations 1 and 2 show the calculations formula to compute the percentage of coagulation performance and turbidity removal.

$$\text{Turbidity Removal} = \frac{\text{Initial Turbidity} - \text{Final Turbidity}}{\text{Initial Turbidity}} \times 100\% \quad \text{Equation(1)}$$

Where  $T_r = \text{Turbidity Removal}$

$T_i = \text{Initial Turbidity}$

$T_f = \text{Final Turbidity}$

$$\text{Coagulation Performance} (\%) = \frac{T_i - T_f}{T_i} \times 100\% \quad \text{Equation(2)}$$

**Table showing the results of Initial parameters of the raw water**

S/N	Parameters	Initial Values
1	Odour	Objectionable
2	Colour	20
3	Taste	Unobjectionable
4	Mouth Feel	Unobjectionable
5	Overall Acceptability	Objectionable
6	pH	7.02

7	Turbidity	450mg/L
8	TDS	550mg/L
9	TSS	2.04g
10	Hardness	19.8mg/L
11	Acidity	8.2mg/L
12	Alkalinity	120mg/L
13	Chloride	40mg/L

**Table 1. Results for Aloe Vera Gel**

Coagulant Dose (g)	Stirring Time (min)	Rate of Settling (min)	Initial Turbidity (NTU)	Final Turbidity (NTU)	Turbidity Removal (NTU)	Coagulation Performance (%)	pH	TDS (mg/L)	TSS (g)
1	30	5	450	25	425	94.4	6.69	180	0.01
2	30	4	450	21	429	95.3	6.72	176	0.01
3	30	4	450	21	429	95.3	6.83	135	0.06
4	30	3	450	17	433	96.2	7.01	117	0.13
5	30	3	450	15	435	96.6	7.10	117	0.16
6	30	2	450	09	441	98.0	7.23	103	0.16

**Table 2. Results for Moringa Seeds Powder**

Coagulant Dose (g)	Stirring Time (min)	Rate of Settling (min)	Initial Turbidity (NTU)	Final Turbidity (NTU)	Turbidity Removal (NTU)	Coagulation Performance (%)	pH	TDS (mg/L)	TSS (g)
1	30	08	450	27	423	94.0	6.90	134	0.01
2	30	08	450	25	425	94.4	7.01	134	0.01
3	30	06	450	23	427	94.8	7.10	122	0.05
4	30	05	450	20	430	95.5	7.21	107	0.10
5	30	05	450	18	432	96.0	7.25	105	0.15
6	30	03	450	13	437	97.1	7.26	103	0.18

**Table 3. Results for Water Melon Seeds Powder**

Coagulant Dose (g)	Stirring Time (min)	Rate of Settling (min)	Initial Turbidity (NTU)	Final Turbidity (NTU)	Turbidity Removal (NTU)	Coagulation Performance (%)	pH	TDS (mg/L)	TSS (g)
1	30	37	450	42	408	90.6	6.80	140	0.20
2	30	36	450	42	408	90.6	6.82	140	0.21
3	30	36	450	41	409	90.8	6.92	138	0.21
4	30	34	450	39	411	91.3	7.01	138	0.25
5	30	29	450	37	413	91.7	7.05	137	0.26
6	30	29	450	37	413	91.7	7.25	132	0.26

**Table 4. Results for Tamarind Seeds Powder**

Coagulant Dose (g)	Stirring Time (min)	Rate of Settling (min)	Initial Turbidity (NTU)	Final Turbidity (NTU)	Turbidity Removal (NTU)	Coagulation Performance (%)	pH	TDS (mg/L)	TSS (g)
1	30	25	450	38	412	91.5	6.92	166	0.30

2	30	25	450	38	412	91.5	7.01	166	0.30
3	30	23	450	37	413	91.7	7.20	162	0.35
4	30	23	450	30	420	93.3	7.22	157	0.32
5	30	20	450	28	422	93.7	7.31	155	0.32
6	30	17	450	28	422	93.7	7.10	155	0.33

**Table5.ResultsforChemicalAnalysis(MoringaSeedsPowder)**

Coagulant Dose (g)	Acidity Removal (mg/L)	Final Acidity (mg/L)	Alkalinity Removal (mg/L)	Final Alkalinity (mg/L)	Chloride Removal (mg/L)	Final Chloride (mg/L)	Hardness Removal (mg/L)	Final Hardness (mg/L)
1	4	4	65	55	23	17	11.5	8.3
2	4	4	65	55	23	17	11.6	8.2
3	5	3	70	50	24	16	12.4	7.4
4	5	3	70	50	27	13	12.4	7.4
5	5.5	2.5	70	50	27	13	12.5	7.3
6	6	3	70	50	29	11	12.2	7.6

**Table6.ResultsforChemicalAnalysis(AloeVeragel)**

Coagulant Dose (g)	Acidity Removal (mg/L)	Final Acidity (mg/L)	Alkalinity Removal (mg/L)	Final Alkalinity (mg/L)	Chloride Removal (mg/L)	Final Chloride (mg/L)	Hardness Removal (mg/L)	Final Hardness (mg/L)
1	3.4	4.6	37.0	63.0	19.0	21.0	10.1	9.7
2	3.6	4.4	36.5	63.5	19.0	21.0	10.5	9.3
3	4.1	3.9	60.9	59.1	19.5	20.5	12.7	7.1
4	5.0	3.0	61.5	58.5	19.6	20.4	12.9	6.9
5	5.1	2.9	64.0	56.0	20.2	19.8	12.9	6.9
6	5.2	2.8	64.1	55.9	22.0	18.0	14.2	5.6

**Table7.ResultsforChemicalAnalysis(TamarindSeedsPowder)**

Coagulant Dose (g)	Acidity Removal (mg/L)	Final Acidity (mg/L)	Alkalinity Removal (mg/L)	Final Alkalinity (mg/L)	Chloride Removal (mg/L)	Final Chloride (mg/L)	Hardness Removal (mg/L)	Final Hardness (mg/L)
1	3.4	4.6	37.0	63.0	19.0	21.0	10.1	9.7
2	3.6	4.4	36.5	63.5	19.0	21.0	10.5	9.3
3	4.1	3.9	60.9	59.1	19.5	20.5	12.7	7.1
4	5.0	3.0	61.5	58.5	19.6	20.4	12.9	6.9
5	5.1	2.9	64.0	56.0	20.2	19.8	12.9	6.9
6	5.2	2.8	64.1	55.9	22.0	18.0	14.2	5.6

**Table8.ResultsforChemicalAnalysis(WaterMelonSeedsPowder)**

Coagulant Dose (g)	Acidity Removal (mg/L)	Final Acidity (mg/L)	Alkalinity Removal (mg/L)	Final Alkalinity (mg/L)	Chloride Removal (mg/L)	Final Chloride (mg/L)	Hardness Removal (mg/L)	Final Hardness (mg/L)

1	2.4	5.6	51.7	68.3	13.7	26.3	9.1	10.7
2	2.2	5.8	51.8	68.2	13.8	26.2	9.6	10.2
3	2.2	5.8	52.7	67.3	14.7	25.3	10.5	9.3
4	1.9	6.1	57.1	62.9	16.0	24.0	10.8	9.0
5	1.8	6.2	57.2	62.8	16.4	23.6	11.2	8.6
6	1.8	6.2	58.8	61.2	16.8	23.2	11.3	8.5

### III. DISCUSSION OF RESULTS

**Table 1** showed the results for stirring time, rate of settling, pH, turbidity removal, coagulation performance, TD Sand TSS for the jar test analysis carried out using Moringa seed powder. The values for the settling time recorded were 8 minutes, 8 minutes, 6 minutes, 5 minutes, 5 minutes and 3 for 1g, 2g, 3g, 4g, 5g and 6g respectively. There was a significant difference between the settling times of 8 minutes and 3 minutes. The values of the pH recorded were: 6.69, 6.70, 6.70, 6.83, 7.10 and 7.25 for 1g, 2g, 3g, 4g, 5g and 6g of the Moringa seed powder respectively. The values of the pH indicated that the mass of the powder increased, the pH increased. This was observed at 1g where 6.69 was recorded and 6g where 7.25 was recorded. There was less significant difference among the values of the pH recorded. This was inline with the results obtained by Gebremicheal et al., (2005) and Aloet al., (2012), who both suggested that pH values of the water samples were not affected by Moringa seed powder, and the values were maintained in the alkaline range.. The pH increased with increased mass of the Moringa seed powder. The action of Moringa seed powder was due to the presence of soluble cationic proteins in these seeds. This suggested that in water, the basic amino acids present in the protein of Moringa will accept a proton from water, resulting in the release of hydroxyl group making the pH of the water alkaline.

The values for the turbidity recorded were: 25 NTU, 21 NTU, 21 NTU, 17 NTU, 15 NTU and 9 NTU for 1g, 2g, 3g, 4g, 5g and 6g respectively. The values of the turbidity removed was calculated by subtracting the values of the final turbidity from the initial turbidity of the raw water which was 450 NTU. The highest turbidity removal value obtained was 441 NTU at 6g and the lowest recorded was 425 NTU at 1g. This indicated that, increased in mass of the powdered seeds, increased the turbidity removal. The values of the final turbidity recorded were with the standard values of turbidity for drinking water given by WHO (2006), where 25 NTU and 5 NTU were given as permissible and acceptable values respectively. This further showed that, Moringa seeds can be adopted for water purification and is likely to reduce cost and also no threat or negative health impact. Also, the percentage of coagulation performance was 98% at 6g and lowest value due to coagulation performance was 94.4% at 1g. This

owed that the Moringa seed powder was effective in removing turbidity of raw water sample.

The values of the TDS recorded were: 180 mg/l, 176 mg/l, 135 mg/l, 117 mg/l, 117 mg/l, and 103 mg/l for 1g, 2g, 3g, 4g, 5g and 6g respectively. The results were in agreement with Mangale et al., (2012), who reported that TDS values were reduced when water was treated with Moringa seed powder. TDS is the amount of both organic and inorganic substances, which can dissolve in a water sample. The initial value of the TDS recorded was 550 mg/l. After the jar test, at 6g, the TDS was 103 mg/l. The higher the turbidity, the higher the TDS and vice versa. Also, the values of the total suspended solids obtained indicated that, as the mass of the powdered seeds increased, the TSS values increased. This was observed at 1g which has 0.01 and 6g which has 0.16. A significant difference of 0.15 g among the values of the TSS was obtained. This was at 6g, there was still some undissolved particles in the water, which lead to an increase in mass of total suspended solids. This was also in agreement with Reena and Harsha, (2019) who recorded a decreased in TSS values as coagulant dose increased using Papaya seed powder and Tamarind Seed powder.

**Table 2** showed the results for rate of settling, pH, turbidity removal, performance of coagulation performance, TD Sand TSS for the jar test analysis carried out using Aloe vera gel. The stirring time was maintained at 30 minutes for each coagulant dose. The values for the settling time recorded were 8 minutes, 8 minutes, 6 minutes, 5 minutes, 5 minutes and 3 minutes for 1g, 2g, 3g, 4g, 5g and 6g respectively. The results showed that the mass of the gel was increased, the settling time decreased. This was observed at 1g of the gel, which took 8 minutes to settle the flocs particles and 6g of the gel took 3 minutes. This showed a significant difference of 5 minutes. The values of the pH recorded were: 6.82, 6.90, 7.00, 7.01, 7.10 and 7.22 for 1g, 2g, 3g, 4g, 5g and 6g respectively. The values of the pH indicated that the mass of the increased, the pH increased slightly, which was observed at 1g with 6.82 and 7.22 respectively. There was less significant difference among the values of the pH recorded. It also showed that, the aloe vera gel has less effect on pH. This was attributed to the organic nature of the aloe vera gel as reported by Abderrezzaq et al., (2021). pH is the degree of acidity or alkalinity of settled water sample.

The values for the turbidity removal obtained were: 423 NTU, 425 NTU, 427 NTU, 430 NTU, 432 NTU and 437 NTU for 1g, 2g, 3g, 4g, 5g and 6g respectively. The value

sof the turbidity removal was calculated by subtracting the values of the final turbidity measured from the initial turbidity of the raw water which was 450 NTU. The highest turbidity removal value obtained was 437 NTU at 6g and the lowest recorded was 423 NTU at 1g. This indicated that, increased in Aloe vera gel increased the turbidity removal. Also, the highest coagulation performance was 97.1% at 6g and pH of 7.22 and lowest coagulation performance was 94% at 1g and pH of 6.82. This showed that pH played crucial role as coagulant are associated with the electric charges according to Hussain et al., (2019). The results recorded were also in agreement with a study conducted by Jaouadi et al., (2020), which showed that Aloe vera gel was an effective coagulant in turbidity reduction and Benalia et al., (2019) which proved turbidity reduction in drinking water by 71.6% and 84.77% from initial value of 13 NTU. The effectiveness of Aloe vera gel as a coagulant was because of the presence of carbonyl function and primary aromatic amine and the presence of NH group in amides as reported by Fatombi et al., (2013). Also, the presence of Glyco-aloe-modinanthrone and tannins were responsible for the coagulation property similar to other natural coagulants as reported by Abonele et al., (2019). The values of the TDS recorded were: 133 mg/l, 133 mg/l, 122 mg/l, 105 mg/l, 105 mg/l and 104 mg/l for 1g, 2g, 3g, 4g, 5g and 6g respectively. TDS is the amount of both organic and inorganic substances, which can dissolve in a water sample. The initial value of the TDS for the raw water recorded was 550 mg/l. After the jartest, the Aloe vera gel at 6g, was able to reduce the initial TDS of 550 to 104 mg/l. This showed that, higher coagulant dose would remove higher TDS. The results of the TDS were in agreement with Mangaleet et al., (2012), who reported a decrease in TDS values as the coagulant dose increased using Moringa seed powder. The higher the turbidity, the higher the TDS and vice versa. Also, the values of the total suspended solids obtained indicated that, as the Aloe vera gel increased, the TSS values increased. This was observed at 1g which has 0.0 g and 6g which has 0.18 g. A significant difference of 0.17 g among the values of the TSS obtained. This was due to the presence of high fibre content in Aloe vera gel, which might not be dissolved completely, and add to the TSS value.

**Table 3** showed the results for rate of settling, pH, turbidity removal, performance of coagulation performance, TDS and TSS for the jartest analysis carried out using Tamarind seed powder. The values for the settling time recorded were 45 minutes, 45 minutes, 43 minutes, 43 minutes, 40 minutes and 38 minutes for 1g, 2g, 3g, 4g, 5g and 6g respectively. The results showed that as the mass of the Tamarind seed powder was increased, the settling time decreased. This was observed at 1g of the, which took 45 minutes to settle the floc particles and 6g of the powder to

ok 38 minutes. This showed a significant difference of 7 minutes. The values of the pH recorded were: 6.92, 7.01, 7.20, 7.20, 7.31 and 7.29 for 1g, 2g, 3g, 4g, 5g and 6g of the respectively. The values of the pH indicated that as the mass of the powdered seeds increased, the pH decreased slightly to weakly alkaline range, which was observed base on the significant difference of 0.91 between the values recorded. It also showed that, this is because, the initial value of the pH recorded for the raw water was 7. which increased slightly to when 1g of the was added, then at 2g the pH value decreased slightly to, then later decreased to an when 5g and 6g was applied. pH is the degree of acidity or alkalinity of settled water sample. This is in agreement with the results recorded by Reena et al., (2019), which shows an increase in pH values as does soft tamarind seed powder increase. The values for the turbidity removal obtained were: 392 NTU, 392 NTU, 403 NTU, 410 NTU, 412 NTU and 412 NTU for 1g, 2g, 3g, 4g, 5g and 6g respectively. The value of the turbidity removal was calculated by subtracting the values of the final turbidity measured from the initial turbidity of the raw water which was 450 NTU. The highest turbidity removal value obtained was 412 NTU at 5g and 6g and the lowest recorded was 392 NTU at 1g. This indicated that, increased in mass of the tamarind seed powder increased the turbidity removal. Also, the percentage of coagulation performance was 91.5% at 6g and lower value of coagulation performance was 87.1% at 1g. This showed that Tamarind seed powder was effective in removing turbidity of raw water sample. This is in agreement with the results obtained by Veena et al., (2019), which showed an increase in turbidity removal, with increase in mass of tamarind seed powder as coagulant. The presence of water soluble proteins with low molecules, amides and hydroxyl groups what makes Tamarind seed to function as not only coagulant, but flocculant as reported by Enrico and Bernard (2019). The active group with negative charges released into the solution, thereby reacting with positive ions or metal ions in the colloidal system. This would help in the formation of fewer flocs and thus disrupting the colloidal system.. The positive charge would react with hydroxyl ions in the suspension to neutralize the pH as reported by Ramadhani et al., (2013). The values of the TDS recorded were: 166 mg/l, 166 mg/l, 162 mg/l, 157 mg/l, 155 mg/l and 155 mg/l for 1g, 2g, 3g, 4g, 5g and 6g respectively. TDS is the amount of both organic and inorganic substances, which can dissolve in a water sample. The initial value of the TDS for the raw water recorded was 580 mg/l. After the jartest, the TDS was reduced to 155 mg/l. The higher the turbidity, the higher the TDS and vice versa. Also, the values of the total suspended solids obtained indicated that, as the increased, the TSS values decrease. This was observed at 1g which has 0.30 and 6g which has 0.33. A significant difference among the values of the TSS was obtained.

**Table 4** showed the results for rate of settling, pH, turbidity removal, performance of coagulation performance, TDS and TSS for the jar test analysis carried out using Watermelon seed powder. The values for the settling time recorded were 37 minutes, 36 minutes, 35 minutes, 35 minutes, 35 minutes and 33 minutes for 1g, 2g, 3g, 4g, 5g and 6g respectively. The results showed that as the mass of the watermelon seed powder was increased, the settling time decreased. This was observed at 1g of the powdered watermelon seeds, which took 37 minutes to settle the flocc particles and 6g which took 33 minutes. This showed a significant difference of 3 minutes. The values of the pH recorded were: 6.80, 6.82, 6.92, 7.00, 7.05 and 6.90 for 1g, 2g, 3g, 4g, 5g and 6g of the watermelon seed powder respectively. The values of the pH indicated that as the mass of the powder increased, the pH decreased slightly towards alkali range, which was observed at 1g with 6.80 and 6g with 6.90. There was a slight significant difference among the values of the pH recorded. It also showed that, has no effect on pH. This is because, the initial value of the pH recorded for the raw water was, decreased slightly to when 6g of the powder was added. pH is the degree of acidity or alkalinity of settled water samples. This is in agreement with the results recorded by Ezeh and Okeke (2017), which showed that increase in watermelon seed powder, increases the pH of the water sample.

The values for the turbidity removal obtained were: 408 NTU, 408 NTU, 409 NTU, 411 NTU, 412 NTU and 412 NTU for 1g, 2g, 3g, 4g, 5g and 6g respectively. The value of the turbidity removal was calculated by subtracting the values of the final turbidity measured from the initial turbidity of the raw water which was 450 NTU. The highest turbidity removal value obtained was 412 NTU at 6g and the lowest recorded was 408 NTU at 1g. This indicates that, increased in dose of the coagulant, increased the turbidity removal. Also, the value of coagulation performance was 91.5% at 6g and the lowest value of coagulation performance was 90.6% at 1g. This showed that was effective in removing turbidity of raw water sample. This agrees with the results recorded by Ezeh and Okeke (2017), where the highest turbidity removal was recorded at higher concentration of dose of the coagulant. The presence of high protein, tannin and mineral contents in the watermelons seeds were responsible for the coagulation properties as reported by Gwana et al., (2014).

The values of the TDS recorded were: 140 mg/l, 140 mg/l, 138 mg/l, 138 mg/l, 137 mg/l and 132 mg/l for 1g, 2g, 3g, 4g, 5g and 6g respectively. TDS is the amount of both organic and inorganic substances, which can dissolve in a water sample. The initial value of the TDS for the raw water recorded was 550 mg/l. At 6g, the TDS was reduced to 132 mg/l from 550 mg/l. The higher the turbidity, the higher the TDS and vice versa. Also, the values of the total suspended solids obtained indicated that, as the Moringa seed powder increased, the TSS values decrease. This was observed at 1g which has 0.20 g and 6g which has 0.26 g.

. A significant difference of 0.1g among the values of the TSS was obtained.

**Table 5 and 6** showed the values recorded for the chemical analysis using Moringa seed powder and Aloevera gel, which showed a decrease in the values as the coagulant dose was increased. This was observed in Moringa seed powder, where 4 mg/L and 3 mg/L were recorded for final acidity, 55 mg/L and 50 mg/L for final alkalinity, 17 mg/L and 11 mg/L for final chloride and 8.3 mg/L and 7.6 mg/L for final hardness at 1g and 6g of the moringa seeds powder respectively. The highest significant difference recorded was 5 mg/L for final alkalinity, and the lowest significant difference recorded was 0.7 mg/L for final hardness. Also, 4.6 mg/L and 2.8 mg/L were recorded for final acidity, 63 mg/L and 55.9 mg/L for final alkalinity, 21 mg/L and 18 mg/L for final chloride and 9.7 mg/L and 5.6 mg/L for final hardness at 1g and 6g each respectively for Aloevera gel coagulant. The highest significant difference of 7.1 mg/L was recorded for alkalinity removal and the lowest significant difference of 1.8 mg/L for final acidity was recorded. This was in agreement with the results obtained by Veena et al., (2019) who reported that values of final acidity, alkalinity and hardness decreased as Moringa and Tamarind seeds coagulants were added.

**Table 7 and 8** showed the results for chemical analysis carried out using watermelon seed powder, showed an increase in values for final acidity, with increase in coagulant dose. This was observed at 1g where 5.6 mg/L was recorded and 6g where 6.2 mg/L and a significant difference of 5.6 mg/L was observed between the highest and the lowest value. But 68.3 mg/L and 61.2 mg/L were recorded for final alkalinity, 26.3 mg/L and 23.2 mg/L for final chloride and 10.7 mg/L and 8.5 mg/L for final hardness, all recorded at 1g and 6g of the coagulant doses respectively. The highest significant difference of 2.3 mg/L was recorded for final hardness.

The values for tamarind seed powder recorded for final acidity showed a slight increase with increased in coagulant dose, then decreased slightly. The values recorded were: 3.6 mg/L, 3.6 mg/L, 3.8 mg/L, 3.8 mg/L, 3.7 mg/L and 3.7 mg/L for 1g, 2g, 3g, 4g, 5g and 6g respectively. Also, the values recorded for final hardness showed similar trend with 7.5 mg/L and 10.4 mg/L were recorded at 1g and 6g respectively. A significant difference of 2.9 mg/L and 0.2 mg/L were recorded for final hardness and final alkalinity. But the values recorded for final alkalinity and final chloride showed that increase in coagulant dose, decreased the values recorded. This was observed at 63 mg/L and 61.8 mg/L and 27 mg/L and 22.6 mg/L for final alkalinity and final chloride respectively. This agrees with the findings of Veena et al., (2019).

#### IV. CONCLUSION

There is a growing interest in the use of natural coagulants derived from plants materials, largely because

e, they are biodegradable and environmentally friendly. The variety of plant-based coagulants includes Aloevera, seeds Moringa, Tamarind and Water Melon as discussed in this research have provided promising coagulation activity and turbidity removal in treating raw water. Therefore, it can be concluded that all the four plants-based materials can remove turbidity in the water and increase coagulation performance, with lesser effect on pH. But aloevera and moringa seed powders showed best potentials and effectiveness in removing turbidity and coagulation performance, then Tamarind seed powder and watermelon seeds.

#### RECOMMENDATIONS

The following recommendations were made:

- Different plant materials should be used like orange seeds, to investigate its potentials as coagulant agent.
- Different plants materials should be blend together to see their potentials and effectiveness.
- Coagulant solutions should be tested to see their effectiveness.
- Further research should be made about natural coagulants of plant-based materials.
- Also, further research should be carried out on how to remove the tastes of the aloevera gel which is bitter and which may affect the taste of the water.

#### REFERENCES

- [1]. AAI Samawi,A.A.,&Shokralla,E.M.(1996).A ninvestigation into an indigenous natural coagulant.Journal of Environmental Science and Health. Part A: Environmental Science and Engineering and Toxicology, 31(8), 1881-1897.
- [2]. APHA,2005. Standard Methods for Examination of Water and Wastewater,20thed. American and Public health association, Washington DC
- [3]. Bhuptawat,H.,Folkard,G.K.,and Chaudhari,S. (2007). Innovative physico-chemical treatment of wastewater incorporating Moringa oleifera seed coagulant.Journal of Hazardous Materials, 142(1), 477-482.
- [4]. Camacho,F.P.,Sousa,V.S.,Bergamasco,R.,and Teixeira,M.R.(2017).The use of Moringa oleifera as a natural coagulant in surface water treatment.Chemical Engineering Journal, 313, 226-237
- [5]. Darge,A.,&Mane,S.J.(2013).Treatment of Industrial Wastewater by Using Banana Peels and Fish Scales. International Journal of Science and Research, 4(7), 600-604.
- [6]. Jiang,J.Q.(2015).The role of coagulation in wastewater treatment.Current Opinion in Chemical Engineering, 8, 36-44.
- [7]. Kansal,S.K.,&Kumari,A.(2014).Potential of M. oleifera for the Treatment of Water and Waste water.Chemical Reviews, 114(9), 4993-5010.
- [8]. Kukic,D.V.,Sciban,M.B.,Prodanovic,J.M.,Tebic,A.N.,and Vasic,M.A.(2015).Extracts of fava bean (*Vicia faba L.*) seeds as natural coagulants. Ecological Engineering, 84, 229-232.
- [9]. Muthuraman,G.,and Sasikala,S.(2014). Removal of turbidity from drinking water using natural coagulants. Journal of Industrial and Engineering Chemistry, 20(4), 1727-1731.
- [10]. Muthuraman,G.,and Sasikala,S.(2014). Removal of turbidity from drinking water using natural coagulants. Journal of Industrial and Engineering Chemistry, 20(4), 1727-1731.
- [11]. Abderrezaq Benalia et al...; Use of aloevera as a norganic coagulant for improving drinking water quality,MDPI , June 2021.
- [12]. Agustin D.,Eric R.,Hermiyanti P.,Diana N.,A.T Narwati: Application of Tamarind seeds Indicas seed extract as bio-coagulant to remove suspended solids and colour. International Journal of Public Health Science Vol 10, June 2021 ( pp 324-329 )
- [13]. Amir Hariz,N.S.Zaidi,Khalida M.,Likewise Wai Loan 2018: A review of effectiveness of natural coagulation process. International Journal of Engineering and Technology 2018 . Vol 7 ( pp 34-37 )
- [14]. Dorghai A.,Moradi M.,Savadpour M.T.,Sharafi K ,: The study of coagulation process in medium turbidity removal from drinking water. Archives of Hygiene Science , 2014 Vol 3. No 4, ( pp 192-200 )
- [15]. Francis K.,Amagloh,Amos 2009:Effectiveness of Moringa seeds as coagulant for water purification. African Journal of Agricultural Research. Vol 4. Feb. 2009 ( pp 119-123 )
- [16]. Muhammad H.,Bin Mohd Lazi 2016:Comparison study of Biopolymer with Alum in the treatment of water,Final Year Project Civil Engineering Department , University of Teknologi Petronas
- [17]. Reena A.,Harsha: Efficiency of Tamarind and Papaya Seed powder as Natural Coagulants. International Research Journal Of Engineering and Technology 2019. Vol 6
- [18]. S.P.Verma et al 2019:Water Treatment By Using Moringa Oleifera and Tamarind Seeds. International Journal of Research in Engineering , Science and Management. Vol 2 May , 2019 issue 5 ( 2581 - 5792 )
- [19]. Vicky K.,Norzila O.,Asharuddin S.,2016: A review of natural coagulants to treat wastewater
- [20]. Hussain,G.; Haydar S. Exploring potential of pearl millet (*Pennisetum glaucum*) and black-

- eyedpea (*Vignaunguiculata* subsp. *unguiculata*) as biocoagulants for water treatment. *Desalination Water Treat.* 2019, 143, 184–191.
- [21]. Fatombi J.K.; Lartiges, B.; Aminou, T.; Barres, O.; Caillet, C. A natural coagulant protein from *Cocos nucifera*: Isolation, characterization, and potential for water purification. *Sep. Purif. Technol.* 2013, 116, 35–40.
- [22]. Saleem, M.; Bachmann, R.T. A Contemporary Review on Plant-Based Coagulants for Applications in Water Treatment. *J. Ind. Eng. Chem.* 2018, 72, 281–297.
- [23]. Bratby, J. *Coagulation and Flocculation within Emphasis on Water and Wastewater Treatment*; Uplands Press Ltd.: Craydon, UK, 1980.
- [24]. Standard Methods for the Examination of Water and Wastewater, 22nd edition. Rice, E.W.; Baird, R.B.; Eaton, A.D.; American Public Health Association (APHA) (Eds.) American Water Works Association (AWWA); Water Environment Federation (WEF): Washington, DC, USA, 2012; ISBN 9780875530130.
- [25]. Benalia, A.; Derbal, K.; Panico, A.; Pirozzi, F. Use of Acorn Leaves as a Natural Coagulant in Drinking Water Treatment Plant. *Water Res.* 2019, 11, 57.
- [26]. Ab Razak N.H., Praveena, S.M., Aris, A., Zand Hashim, Z. (2015). Drinking water studies: A review on heavy metal application of biomarker and health risk assessment (a special focus in Malaysia). *Journal of Epidemiology and Global Health*, 5 (4), 297-310. doi: <http://dx.doi.org/10.1016/j.jegh.2015.04.003>
- [27]. Bratby, J. (2006). *Coagulation and flocculation in water and wastewater treatment*; IWA publishing.
- [28]. Kulinkina, A.V., Kosinski, K.C., Liss, A., Adjei, M.N., Ayamgah, G.A., Webb, P., ..., Naumova, E.N. (2016). Piped water consumption in Ghana: A case study of temporal and spatial patterns of clean water demand relative to alternative water sources in rural small towns. *Science of The Total Environment*, 559, 291-301. doi: <http://dx.doi.org/10.1016/j.scitotenv.2016.03.148>